## JOURNAL OF THE

## LABORATORY NOTES ON THE STABILIZATION OF FLUIDEXTRACT OF ERGOT.\*

# BY ELMER H. STUART AND FRANCIS E. BIBBINS.

In spite of the large number of published articles relating to the deterioration of Fluidextract of Ergot, the problem of a satisfactory method of stabilizing this preparation has not yet been solved. Because of this fact the laboratory notes herein reviewed are submitted with the hope that they may be of value to other workers interested in this problem. The authors wish to point out that many of the ideas are not original but have been gathered from the literature as suggestions worthy of investigation.

The various assay results reported in this paper were carried out according to one of three well-known methods of determining potency of ergot and its preparations. These methods were, *first*, a modification of the Broom and Clark method (1), *second*, the Cock's Comb method of the U. S. P. X and *third*, a modification of Smith's Colorimetric method (2).

Swanson in 1929 (3) pointed out that the control of the  $p_{\rm H}$  of Fluidextract of Ergot is important. Our data, however, would indicate that the problem is more complicated than that of adjusting the  $p_{\rm H}$ , by the addition of acid to the Fluidextract.

The various methods for determining  $p_{\rm H}$  were investigated for a reliable method applicable to Fluidextract of Ergot and one which could be readily checked by laboratory chemists. The hydrogen electrode was discarded because the results are questionable in the presence of alcohol. The removal of the alcohol before determining the  $p_{\rm H}$  so altered the Fluidextract that the figures obtained were of little value. The quinhydrone electrode [Coons (6)] gives  $p_{\rm H}$  values which are readily duplicated when the following method is observed:

Add 0.1 Gm. quinhydrone (Eastman) to 5-cc. Fluidextract of Ergot. Adjust the temperature to  $25^{\circ}$  C. and read after allowing two or more minutes for the electrode to reach equilibrium. Before using the electrode it should be cleaned as Coons suggests by boiling for five minutes in 50% nitric acid solution, rinsing with distilled water, followed by boiling for five minutes in 10% sodium bisulphite solution and again rinsing with distilled water.

It has been the observation of the authors that some Fluidextracts of Ergot appear to be relatively stable while others made by the same method from another lot of ergot deteriorate rapidly. It is our opinion that some lots of crude ergot contain something that stabilizes the Fluidextract while other lots do not contain sufficient of this material to exercise the stabilizing influence for any appreciable time. In an attempt to answer this question the ash was determined from a number of Fluidextracts of Ergot in order to see if this factor had any relation to stability. The ash varied from 1.33 Gm. to 2.09 Gm. per 100 cc., but showed no relation to stability. The iron in the ash was next determined and was found to range from a trace up to 0.25 Gm. per 100 cc. calculated to ferric oxide. Again there was no relation to stability. Fluidextract of Ergot also contains traces of copper, aluminum and nickel. As a further check on the influence of metals, the percolation was carried out in glass, iron and monel percolators (see Table I). The results were negative.

<sup>\*</sup> Scientific Section, A. PH. A., Washington meeting, 1934.

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Since some Fluidextracts of Ergot are more stable than others, it occurred to the authors that perhaps this condition might be due to variation in the proportion of rye grains contained in different lots of ergot. This was checked by adding to the Fluidextract an extract of rye prepared in the same manner as the Fluidextract of Ergot. The result obtained was negative (see Table II).

					$\mathcal{P}_{\mathbf{H}}$	Assay.						
Davis		6 1	A 1 1-1-		after	Bre		nd Cla	rk.	ູບ	. S. P. J	
Drug No.		Sample No.	Additions per 500 Cc.	∲н.	Aging. 6 Mos.	Mo.	4 Mos.	8 Mos.	۱ ۲۲.	I Mo.	3 Mos.	8 Mos.
Α		P-22596		4.5	4.75		140	90	55		120	<b>25</b>
	Glass	P-22596 B	7.5 cc. 36% HC1	3.1				100				
	percolator	P-22596 D	15.0	2.2			160				115	
		P-22596 F	8.2	3.0	3.3	330	125	100	108	120	100	50
		P-22597		4.6			125					
	Monel	P-22597 B	7.5 cc. 36% HC1	3.1			150	100	97	300	300-	<b>25</b>
										320	320	
	percolator	P-22597 C	8.25	3.0				100				100
		P-22597 D	10.0	2.9		160						
		P-22597 E	15.0	2.35			150				120	
в		P-22733	· · · · · · · · · · · · · · · · · · ·	4.7			40		30		25	
		P-22733 C	8.25 cc. 36% HCl	3.1				15				15
	Glass percolator	P-22733 D	10.0	2.9	••	100	50	20	30	100	50 4 Mos.	2 <b>0</b>
		P-22733 F	20.0	2.15	•••		$^{25}$				33	

#### TABLE I.—FLUIDEXTRACT OF ERGOT.

### TABLE II.—FLUIDEXTRACT OF ERGOT.

Drug No.	Sample No.	Additions per 500 Cc.	<i>∲</i> <sub>H.</sub>	₽ <sub>H</sub> after Aging.	Br 1 Mo.	coom and 4 Mos.	l Clark. 8 Mos.	Assa 1 Yr.	<sup>ay,</sup> U 1 Mo.	. S. P. 1 4 Mos.	X. 8 Mos.
		per 500 Cc.			1410.			11.	1010.		
С	P22792		4.6	4.8		120	75			200	50
	P22792 C	8.25 cc. 36% HC1	3.0	3.35	125	200	100	105	130	120	100
	P22792 E	15.0	2.2	2.7		100		80		50	
	P22792 F	20.0	1.65	· ·			100				100
D	P23803		3.7		100	100			100		
	P23804	3.6 cc. 36% HCl	3.0		100	100-				Colori	metric
					110	110			120	8 M	
	P23814	3.6 cc. 36% HC1							8 Mos.		
		1% milk sugar	3.0			30	35	25	40	1	5
	P23815	3.6 cc. 36% HCl									
		1% cane sugar	3.0			66 <sup>2</sup> /3	51	25	40	3	5
	P23816	3.6 cc. 36% HCl									
		2.5% dextrose	3.0			100	100	62	80	e	5
	P24151	3.6 cc. 36% HC1									
		2.5% rye extract	3.0					65			
	P24152	3.6 cc. 36% HC1									
		10 Gm. Hydroqui-									
		none	3.0					<b>94</b>			

The addition of cane sugar, milk sugar and dextrose to Fluidextract of Ergot resulted in increased deterioration (see Tables II and III). As an additional check on this point, two relatively stable and two unstable Fluidextracts of Ergot were tested for their reducing action of Fehling's solution (U. S. P. X, page 497) as follows:

Five-cc. Fluidextract of Ergot were evaporated so as to remove the alcohol, then 15 cc. each of Fehling's solution A and B added. The mixture was diluted to 100 cc., then boiled for two minutes and filtered through a tared Gooch crucible. The precipitate was washed with distilled water, followed by alcohol and by other and then dried thirty minutes at 100° C. The blank was run in the same way or itting Fehling's solution A (copper sulphate solution).

Sample No. 1 (Stable)	1 cc. reduced 0.0214 Gm. CuO.
Sample No. 2 (Unstable)	1 cc. reduced 0.0116 Gm. CuO.
Sample No. 3 (Stable)	1 cc. reduced 0.0157 Gm. CuO.
Sample No. 4 (Unstable)	1 cc. reduced 0.0228 Gm. CuO.

## TABLE III.---FLUIDEXTRACT OF ERGOT.

		TABLE IIIFLU	IDEXTRACT OF E	RGOT.				
						Assay		
Drug No.	Sample No,	Method of Percolation. U. S. P. X Type A. Fluidextract.	Additions per 500 Cc.	₽ <sub>н.</sub>	Broom a 1 Mo.	und Clark 2 Mos.	. Colo 1 Mo.	rimetric. 2 Mos.
	P24460	50% alcohol	6 cc. 36% HCl	3.0	110		100	
Е	P24461	50% alcohol plus 9.1 cc. 36% HCl per 500 Gm.		4.2	100		110	
	P24462	66 <sup>2</sup> / <sub>3</sub> % alcohol	3.5 cc. 36% HCl	3.0	108		100	
	P24463	66 <sup>2</sup> /3% alcohol plus		4.2	133		125	
		9.1 cc. 36% HCl per 500 Gm						
	P24464	75% alcohol	2.5 cc. 36% HC1	3.0	106	110	110	
	P24465	75% alcohol plus 9.1 cc.						
		36% HCl per 500 Gm.		4.2	140		150	
	P24466	85% alcohol	3.0 cc. 36% HC1	3.0	130	90	140	
	P24467	85% alcohol plus 9.1 cc.						
		36% HC1 per 500 Gm.		<b>4</b> .2	150		160	
	P24523	95% alcohol plus 9.1 cc.						
		36% HCl per 500 Gm.		4.4	177		120	
F	P24514	50% alcohol plus 9.1 cc.		4.6		167		130
	P24515	36% HCl per 500 Gm.	2.5% dextrose			109		95
	P24516		2.5% sodium					
			thiosulphate			80		100
	P24517	85% alcohol plus 9.1 cc.		4.8		200	200	<b>20</b>
	P24518	36% HCl per 500 Gm.	2.5% dextrose			18		20
	P24519		2.5% sodium					
			thiosulphate			30		20
	P24520	95% alcohol plus 9.1 cc.			135	115	140	100
	P24521	36% HCl per 500 Gm.	2.5% dextrose			62		80
	P24522		2.5% sodium					
			thiosulphate			116		93

# TABLE IV.—FLUIDEXTRACT OF ERGOT, U. S. P. X.

# $p_{\rm H}$ Previously Adjusted to 3.0 with HCl.

		$p_{ m H}$ Previously Adjusted to	3.0 with	HCl.			
Lot Fluid- extract of Ergot.		Additions.		om and C after Agin 4 Mos.		U. S. P. X. C	colorimetric. 1 Mo.
1	P24269	• • • • • • • • • • • • • • • • • • • •	100	90	84	97	100
	P24270	Aerated 48 hours	100			114	116
	P24271	0.5% petroleum benzin then					
		aerated 48 hours	100	38			96
							4 Mos.
	P24272	<sup>1</sup> / <sub>4</sub> full bottle		38			45
							1 Mo.
<b>2</b>	P24274		120	1 <b>2</b> 0	123		1 <b>2</b> 0
	P24275	Aerated 48 hours	90				110
	P24276	0.5% petroleum benzin then					
		aerated 48 hours	100			120	100
	P24277	<sup>1</sup> / <sub>4</sub> full bottle		58			60
				2 Mos.			
1	P24716	· · · · · · · · · · · · · · · · · · ·		120			
	P24718	0.65% borneol		72.5			
	P24719	0.65% benzaldehyde		40			
	P24720	10% acetone		44			
	P24721	0.65% hydroxy methyl ane-					
		thol		74			
	P24722	0.65% NaH <sub>2</sub> PO <sub>4</sub>		54			
	P24756	0.1% quinaldine		71			

Various substances that might function as antioxidants and inhibitors were added to the Fluidextract of Ergot, namely, hypophosphorous acid, linseed oil, vitamin A concentrate, hydroquinone, carotene, sodium thiosulphate, sodium hydrosulphite, ferrous sulphate, ergosterol, cholestrin, borneol, benzaldehyde, acetone, quinaldine and hydroxy methyl anethol. Of the foregoing compounds only the first four appeared to show any stabilizing influence on Fluidextract of Ergot (see Tables II, III, IV and V).

Lot Fluidextract of Ergot.	Sample No.	Additions.	Assay. Broom and Clark after 3 Wks. at 50° C.
3	P26417	0.025% carotene	73
	P26418	0.025% vitamin A concentrate	89
	P26419		73
	P26625	0.5% ergosterol from ergot	55
	P26626	•••••••••••••••••••••••••••••••••••••••	51
	P26627	0.5% cholestrin	<b>46</b>
	P26628	0.5% vitamin A concentrate	95
4	P26634	0.5% ergosterol from ergot	63
	P26635	0.5% cholestrin	55
	P26636	0.5% vitamin A concentrate	50
	P26637	• • • • • • • • • • • • • • • • • • • •	41
5	P26910	0.5% vitamin A concentrate	40
	P26911	0.5% cod liver oil	45
	P26912	0.5% alcoholic extract cod liver oil	49
	P26913		25
	P27028	0.5% cod liver oil	45
	P27029	0.5% linseed oil	60
	P27030	0.5% liquid petrolatum	50
	P27031	0.5% cottonseed oil	49
	P27032	0.5% ergot oil (petroleum benzin)	38
	P27033	• • • • • • • • • • • • • • • • • • • •	38
6	P27022	0.5% cod liver oil	50
	P27023	0.5% linseed oil	60
	P27024	0.5% liquid petrolatum	40
	P27025	0.5% cottonseed oil	47
	P27026	0.5% ergot oil	44
	P27027		50

# TABLE V.—FLUIDEXTRACT OF ERGOT, U. S. P. X. $p_{\rm H}$ Previously Adjusted to 3.0 with HCl.

The method of the British Pharmacopœia, 1932, requires extraction of the ergot with fifty per cent alcohol containing one per cent of tartaric acid. The Fluidextract of Ergot made by this method (see Table VI) deteriorated within six months. This is about the same rate of deterioration as is observed when the U. S. P. X method of preparation is used. The addition of tartaric or of hydrochloric acids to the Fluidextract did not give a stable product as is shown by Tables VI, VII and VIII.

In Tables III, VI and IX the method of percolation was changed from the fractional percolation procedure of the U. S. P. X to the single percolation process which is described on page 159 of the Pharmacopœia as process Type A. This method of percolation yielded a Fluidextract of Ergot which is as active as the regular U. S. P. X method when made from the same lot of drug (compare Table VI).

Wokes and Elphick (4) found that defatting ergot increased the efficiency of the extraction with either neutral or acidified alcohol. Our results as shown in Table IX do not support this statement. As a matter of fact we have prepared a standard Fluidextract of Ergot by percolating the whole drug without having applied any preliminary treatment. Our attempt to answer the question as to whether the stability of the Fluidextract is influenced by the incomplete removal of the petroleum benzin from the drug before percolation is shown in Table IV. The addition of petroleum benzin to the Fluidextract apparently had no effect on the stability.

		IADDE V.	I. I DOIDBAIKACI	OF L						
					Bro	om an		ays. k.	u. s.	Р. Х.
Drug	Sample	Method of	Additions		1	4	6	1	3	6
No.	No.	Percolation.	per 500 Cc.	⊅н	Mo.	Mos.	Mos.	Yr.	Mos.	Mos.
в	P22875 A	The 1932 British								
		Pharmacopœia	3.7 Gm. tartaric	4.0		50			<b>25</b>	
	P22875 C		11.1 Gm. tartaric	3.1			50			80
	P22875 D	(1% tartaric in 50%	12.95 Gm.	3.0	160	100	<b>25</b>	25	50	33
	P22876 B	alcohol)	9.0 cc. 36% HCl	3.0			25	65	25	33
Е	P23454	U. S. P. X			50				33	
	P23454 D	Iron fillings mixed								
		with ergot	20 Gm. tartaric				70			
	P23453	U. S. P. X		5.8	35				33	
	P23453 B	U. S. P. X	7.5 cc. 50% H <sub>3</sub> PO <sub>2</sub>	3.0			90			
	P23452	U. S. P. X		5.4	100			40	<b>70</b>	
	P23452 B	Iron sulphate dried	3.0 cc. 36% HC1					70		
	P23452 D	powder mixed with ergot. 4 Gm. per 500 Gm.	20 Gm. tartaric				110			
	P23455 A P23455 B	U. S. P. X type A Fluidextract 50%	20 Gm. tartaric				100			
		alcohol	5.0 Gm. Fe <sub>2</sub> (SO <sub>4</sub> );				105			

TABLE VI.—FLUIDEXTRACT OF ERGOT.

TABLE	VII.—Fluidextract	OF	Ergot.
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		INDLE VII.	1 001	DUAIRA		Jr DAG	<i>.</i>				
								Assa	ys.		
				₽ <sub>H</sub>		Broom an	d Clar	k.	U.	S. P. 3	x.
Drug	Sample	Additions		after	1	4	9	2	2	5	9
No.	No.	per 500 Cc.	⊅н.	Aging.	Mo.	Mos.	Mos.	Yrs.	Mos.	Mos.	Mos
в	P27732 B	2.5 cc. 36% HCl	4.6			331/3			100	50	25
				6 Mos.							
	P27732 D	7.5	3.0	3.2	40	331/3	20	30	100	50	25
	P27732 E	10.0	2.5				15				
	P27732 F	12.5	1.9			18	12			25	
	P27732 M	5.55 Gm. tartaric acid	4.45			331/2				30	
	P27732 N	7.40	4.3				20				25
	P27732 P	11.10	3.5				12				25
	P27732 Q	12.95	3.3			50				25	
с	P22791 A	3.70 Gm. tartaric	4.7			50				<b>6</b> 2	
	P22791 B	7.40	4.2				50				62
				8 Mos.							
	P22791 E	14.80	3.0	3.25			50	118			63
	P22791 F	16.65	3.0	3.20		100	100	100		120	100 - 120

In Table IV the effect of air upon Fluidextract of Ergot was observed by bubbling through the sample, for 48 hours, a current of air previously saturated with the menstruum by passing the air through diluted alcohol. No immediate loss of activity was observed, although, on aging the samples, the deterioration progressed more rapidly.

In 1930 Thompson (7) recommended that Fluidextract of Ergot be distributed in completely filled bottles in order to prevent undue exposure to air. Our findings (see Table IV) agree with his conclusions, since deterioration progressed more rapidly when stored in bottles that were one-fourth full, Nov. 1934

Our results shown in Table III agree with Linnell and Randle (5) in that an acid alcohol menstruum is more efficient than alcohol alone. The table also indicates that the higher percentage alcohol extracts the activity more completely than does alcohol of lower concentration. All samples, however, deteriorated upon aging.

### TABLE VIII.-FLUIDEXTRACT OF ERGOT.

				₽ <sub>H</sub>						
<b>T</b>	0 1	Additions		after	п.			ssays.		-
Drug No.	Sample No.	per 500 Cc.	<b>⊅</b> н.	Aging. 9 Mos.		oom an 9 Mos.		2 Mos.	U. S. P. 2 4 Mos.	9 Mos.
Α	P-22595 A		4.7		100	100		100	80	<b>25</b>
	P-22595 D	7.5 cc. 36% HC1	2.95	3.05	160	66	69	115-130	70	25
	P-22595 F	12.5 cc. 36% HCl	2.2		110	100			40 - 50	25
	P-22595 I	6.0 cc. 36% HCl	3.1		100	100				50
	P-22595 K	1.85 Gm. tartaric acid	4.45		130	90			5 Mos.	<b>25</b>
									25	
	P-22595 P	11.10 Gm. tartaric acid	3.15			50				
	P-22595 Q	12.95 Gm. tartaric acid	2.95		160				33	
С	P-22790		6.0							<b>25</b>
	P-22790 A	2.5 cc. 36% HC1	4.7		120	100	66²/3		62.5	
	P-22790 D	7.5	3.0			100	$66^{2}/_{3}$	80	100	100-110
	P-22790 E	10.0	2.5				331/3			50
	P-22790 F	12.5	2.0			50			5070	

#### TABLE IX.—FLUIDEXTRACT OF ERGOT.

						As	says.	
D	0	36-411-6	Additions		Broom a			S. P. X.
Drug No.	Sample No.	Method of Percolation.	per 500 Cc.	<b>⊅</b> <sub>H.</sub>	1 Mo.	5 Mos.	1 Yr., 4 Mos.	1 Mo,
Е	P23500	U. S. P. X type A Fluid-		4.6			85	
	P23500 A	extract menstruum—50%	5.0 cc. H <sub>3</sub> PO <sub>2</sub>	3.1	60	150	150	100
	P23500 B	alcohol plus 7 cc. 50%	7.5 cc. H <sub>3</sub> PO <sub>2</sub>				155	
	P23500 C	H₃PO₂ per 500 Gm. ergot	5 Gm. Fe(SO4)3			120	200	
	P23500 D		20 Gm. tartaric		90-100	150	112	100
	P23500 E		3.75 cc. HCl 36%	3.0		140	135	100
F	P24613	U. S. P. X except used					Colori	metric.
		whole ergot not defatted		4.3	185		1	60
	P24614	U. S. P. X except ergot						
		not defatted		4.65	180		1:	20
	P24615	U. S. P. X except used						
		whole ergot		4.45	130		1	30
	P24616	U. S. P. X		4.75	113		14	<b>5</b> 0
	P24710	U. S. P. X except used						
		whole ergot not defatted	· · · · · · · · · · · · · · · · · · ·	4.3	180		2 M	os.
							10	30

## CONCLUSIONS.

1. Fluidextract of Ergot is not stabilized by adjusting the  $p_{\rm H}$  by means of acids.

2. A reliable method for determining the  $p_{\rm H}$  of Fluidextract of Ergot is outlined.

3. Some lots of crude ergot contain something which tends to stabilize the Fluidextract more than others.

4. The addition of rye extract to the Fluidextract of Ergot apparently does not influence the stabilization.

5. The addition of sugars increases the deterioration of Fluidextract of Ergot.

6. The power of Fluidextract of Ergot to reduce Fehling's solution bears no relation to stability.

7. Hypophosphorous acid linseed oil, vitamin A concentrate and hydroquinone favorably influence the stability of Fluidextract of Ergot.

8. Fluidextract of Ergot made by the method of the British Pharmacopœia deteriorates the same as that made by the U. S. P. X method.

9. The type of percolation is not important.

10. Defatting the drug before percolation is not important.

11. The activity of ergot is extracted more completely by acid alcohol than by neutral alcohol.

12. Higher percentage alcohol is more efficient for percolation of ergot than low percentage alcohol.

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THE LILLY CONTROL LABORATORIES, INDIANAPOLIS, INDIANA.

## THE TESTING OF ERGOT.

## BY HOWARD H. CROSBIE.

In the course of investigating the breakdown rate of liquid preparations of ergot, we have in this laboratory been using all three usual methods of testing, i. e., the Broome-Clarke rabbit uterus method, the Allport-Cocking color reaction, and the Cock's Comb method, with a distressing want of correlation, driving one to the verge of despair. We have experimented with a photographic modification of the Cock's Comb reaction that we think it worth drawing to the attention of other workers.

The method is to photograph the bird, before injection, by means of an appropriate light filter and red sensitive plates so that blue registers as black and red registers as white. The bird is then injected and after  $1^{1}/_{4}$  hours is again photographed on the same plate, consequently the two photographs get the same development. The resultant prints although not necessarily good pictures of birds do pick up differences that are not visible to the unaided eye.

Before making an assay, one prepares two pairs of reference prints, one pair with a dose of some standard (in this case Ergotoxine ethanesulphonate solution 1/2 mg. per cc.) of such size as to produce a minimum effect as in Fig. 1. Another reference photograph is made of the same bird with a larger dose and more pronounced effect as in Fig. 3. In assaying a sample marked "A" a first trial was made on the assumption that it was probably over-strength and a lesser dose of "A" was given than had been given to the same bird in Fig. 1 with the result shown